

## Biomass production and biochemical variability of marine microalgae *Isochrysis galbana* in relation to light intensity

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### Abstract

*Isochrysis galbana* is a marine microalga that has a great potential as a source of animal feed, human nutrition and biofuel. Light intensity is one of the key limiting factors for the growth of dense culture of photosynthetic microalgae both in indoor and outdoor culture. In this study, we investigated the effect of different light intensity (35, 125 and 275  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ ) on the biomass production and its biochemical composition (protein carbohydrate, lipid and pigments). Growth is calculated daily by measuring optical density at 710nm and dry weight of filtered cells was incinerated at 550°C in a muffle oven. The biochemical components protein, lipid, carbohydrate and pigment were measured by Lowry, Bligh and Dryer, Dubois and Strickland and Parsons Method respectively. In this study, reported culture (LL) grown under low light have maximum amount of protein content ( $44.73 \pm 0.50\%$  DCW) and chlorophyll a ( $4.67 \pm 0.28 \text{ mg/L}$ ) whereas culture (HL) grown under high light intensity were highest in the lipids contents ( $30.66 \pm 0.30\%$  of DCW) and carbohydrates ( $16.06 \pm 0.70\%$  of DCW). Maximum accumulation of total carotenoids ( $4.89 \pm 0.25 \text{ mg/L}$ ) reported in culture (HL) grown under high light and with decreased light carotenoids decreases. In conclusion result reported that varied light intensity have a significant impact on growth, dry weight and biochemical composition of *Isochrysis galbana* which have wide application in aquaculture, nutraceutical, pharmaceutical and biofuel industry.

**Keywords:** *Isochrysis galbana*, light intensity, biochemical composition, pigments

### 1. Introduction

*Isochrysis galbana* is one of the common marine algae have been used in mariculture to feeds for bivalves & larva of fish, crustaceans, mollusks in aquaculture [1]. Its high nutritional value, small size, higher growth rate and simpler growth requirement, attracts the nutritionist and biotechnologist. They have the ability to grow in extreme environmental condition and in order to adopt these conditions they accumulates varieties of bioactive compounds that can be used in bioremediation [2], biofuel [3], biofertilizer [4], human food [5], animal feed [6], and pharmaceutical industry [7]. In order to have microalgae with high nutritional value various researches have been conducted around world. The nutritional value of microalgae relates to its biochemical composition which varies with growth phase and culture conditions like light, temperature, nitrogen concentration or salinity [8]. Among various environmental factors, light is one of the key factors that control the various physiological processes and thereby their growth and biomass production. Quantity and quality of light determines the amount of energy available to photosynthetic organisms to conduct their metabolic activities [9]. Therefore, it is the most important factor affecting the photosynthesis kinetics productivity of microalgae [10]. In large scale cultivation, dense culture of microalgae limits the penetration of light which affects the photosynthesis and growth of microalgae. Previous studies have shown that under stress they alter their metabolic pathway from growth promoting to energy saving pathway [10]. Thereby it changes their biomass yield, growth rates and biochemical composition which have diverse application in the aquaculture,

pharmaceutical, nutraceutical, cosmetic, and biofuel industry. Variation of irradiance and depletion of nutrients is the critical event occurs in both indoor and outdoor cultivation which affects biomass production and its composition. The objective of this study is to point out how change in light intensity in batch culture affects the growth and biochemical composition of microalgae. As microalgae is valuable bio-resources for animal feed, human nutrition, nutraceutical and biofuel industry on the world market, it raises the need of improved knowledge of its composition, growth and chemical variability for higher production of particular valuable metabolites with high mass productivity. Therefore it is important to investigate the effect of light limitation on growth and biochemical composition of microalgae.

### 2. Materials and Methods

#### 2.1 Algal culture

The starter culture of marine microalgae *Isochrysis galbana* was obtained from Center of Marine and Fishery Research Institute (CMFRI) Kochi, India.

#### 2.2 Growth condition

The experiment was carried out in 1litre conical flask containing 200-300ml of growth media. The growth media contained autoclaved filtered natural seawater, supplemented with f/2 culture medium [11]. The f/2 culture medium was composed of  $75 \text{ mg L}^{-1} \text{ NaNO}_3$ ,  $5 \text{ mg L}^{-1} \text{ NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$ ,  $1 \text{ ml L}^{-1}$  trace metal solution ( $3.15 \text{ mg L}^{-1} \text{ FeCl}_3 \cdot 6\text{H}_2\text{O}$ ,  $4.36 \text{ mg L}^{-1} \text{ Na}_2\text{EDTA}$ ,  $0.0098 \text{ mg L}^{-1} \text{ CuSO}_4 \cdot 5\text{H}_2\text{O}$ ,  $0.0063 \text{ mg L}^{-1} \text{ Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ ,  $0.022 \text{ mg L}^{-1} \text{ ZnSO}_4 \cdot 7\text{H}_2\text{O}$ ,  $0.01 \text{ mg L}^{-1}$

CoCl<sub>2</sub>·6H<sub>2</sub>O, 0.18 mgL<sup>-1</sup> MnCl<sub>2</sub>·4H<sub>2</sub>O) and 0.5 ml/L of vitamin solution (0.001 mg L<sup>-1</sup> vitamin B12, 0.2 mgL<sup>-1</sup> vitamin B<sub>1</sub>, 0.001 mg biotin<sup>-1</sup>) were added. In the laboratory, *Isochrysis galbana* grown in a batch culture at temperature of 22-25 °C, pH 8-8.2 and under different light intensity: 35 μmol photon m<sup>-2</sup> s<sup>-1</sup> (low), 125 μmol photon m<sup>-2</sup> s<sup>-1</sup> (medium), and 275 μmol photon m<sup>-2</sup> s<sup>-1</sup> (high), provided by white fluorescent tube for 24:0 h light-dark period. All the glassware and media were always sterilized prior to inoculation. All the experiments were carried out after 5<sup>th</sup> of cultivation and in triplicates. For this biomass was harvested after 5 days by centrifugation at 10,000 rpm for 10 min, washed twice with distilled water.

### 2.3 Measurement of Algal growth

The growth of *Isochrysis galbana* is calculated by measuring optical density (O.D.) and dry weight. Optical density was measured daily at 710nm by UV/visible spectrophotometer. For dry weight measurements, 50ml of culture samples were centrifuged at 5000 rpm for 5 min. After rinsing twice with distilled water, the pellets were dried 6hr in an oven at 110°C and then cooled down in desiccators before weighing. The difference between the initial and final weight were taken as the dry weight of algal biomass (mg/l).

### 2.4 Biochemical Analysis

The crude protein was determined by modified Lowery method [12]. The absorbance of the sample was taken at 650nm and the concentration was determined using standard curve: Total Protein Content = wt. of protein (from curve) X 100/ dry cell mass (mg). The content of carbohydrate is estimated by the modified by the phenol-sulfuric acid method of Dubois [13]. The optical density of the sample was determined against the blank at 490 nm in a UV-visible spectrophotometer. Carbohydrate Content (%) = wt. of carbohydrate (from

Glucose standard curve) X 100/ dry cell mass (g) and total lipid contents were analyzed gravimetrically after extraction with chloroform-methanol (2:1) modified by Bligh and Dyer [14]. Pigments were extracted in acetone (90 %) at 4 °C overnight and measured by spectrophotometric methods [15].

### 2.5 Data Analysis/Statistical Analysis

All experiments were done with three replicates and data represent the means ± SD. They were analyzed by one-way ANOVA and significant differences between treatments were tested using Duncan's multiple range test (DMRT). *P*-values <0.05 were considered significant. Statistical analyses were carried out using Statistical Package for the Social Sciences (SPSS) 10.0.

## 3. Results

### 3.1 Growth pattern

Light depletion is one of the key limiting factor affecting the growth and biomass production. To evaluate the effect of light intensity on biomass productivity absorbance at 710nm was measured. Figure 1 illustrated the growth curve of *Isochrysis galbana* under different intensities of light. It elicited that growth was significantly (>0.05) higher in culture under high light intensity. Growth measured in term of dry weight was shown in figure 2. There was a significant difference (>0.05) between all the cultures under different light intensity. The culture (ML) grown under medium light attained a highest dry weight (0.97±0.028g/L) followed by culture HL (0.83±0.042g/L). At end of cultivation (5day), the dry weight increased from 0.6 mg/L to 0.97 mg/L with increased light intensity from 35 μmol photons m<sup>-2</sup> s<sup>-1</sup> to 125 μmol photons m<sup>-2</sup> s<sup>-1</sup> but further increase of light intensity from 125 to 275 μmol photons m<sup>-2</sup> s<sup>-1</sup> decreased dry weight from 0.97mg/L to 0.83mg/L had observed.

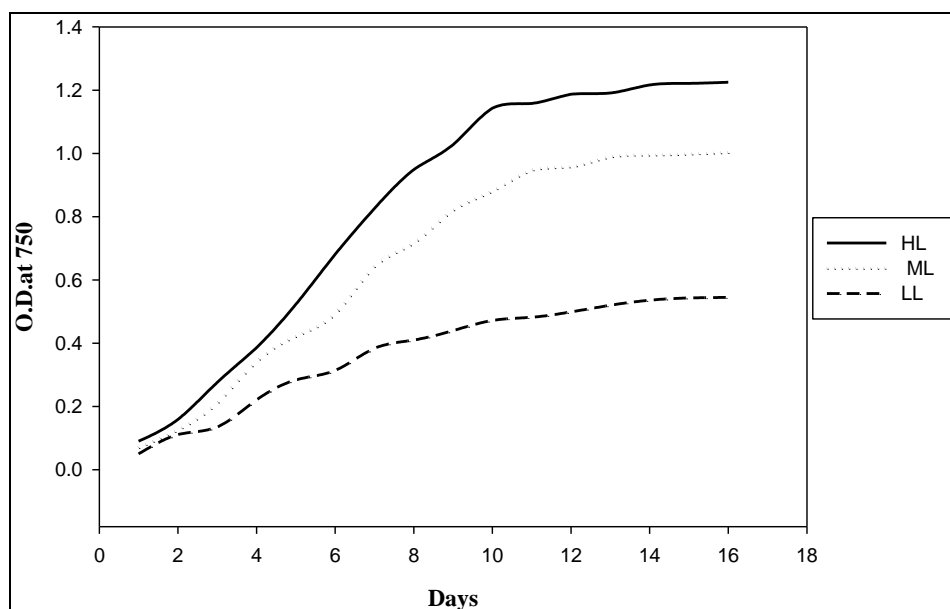


Fig 1: Growth pattern of *Isochrysis galbana* under different light condition

a) high light intensity of 35 μmol photon m<sup>-2</sup> s<sup>-1</sup>, b) medium light intensity of 125 μmol photon m<sup>-2</sup> s<sup>-1</sup>, c)

low light intensity of 275 μmol photon m<sup>-2</sup> s<sup>-1</sup>.

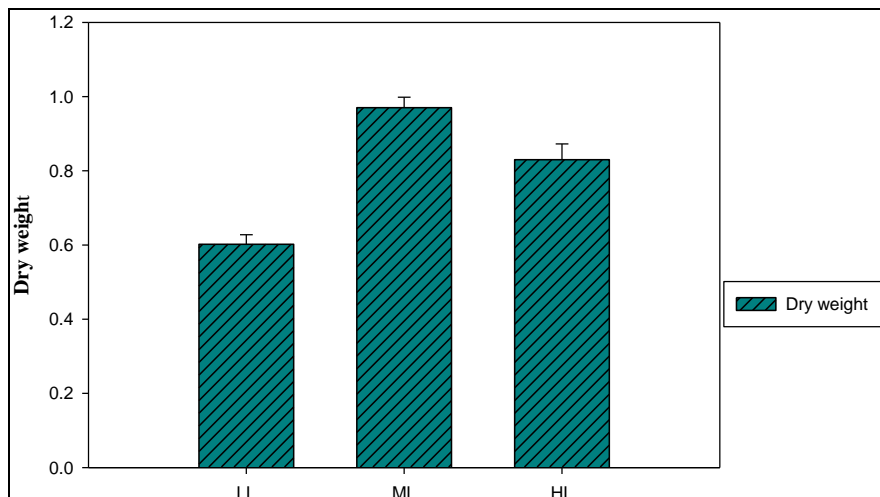


Fig 2: Effect of light on biomass yield of *I. galbana*.

### 3.2 Biochemical composition & Pigment

In this study, culture were grown in three light regime (LL,ML,HL) and its effect on biochemical composition and pigment content were shown in table1. It had reported that in each light regime, all biochemical parameters were significant different ( $p < 0.05$ ). The variations of the determined biochemical compositions during the cultivation of *I. galbana* under different light intensity are illustrated in figure 3. With increased light intensity, carbohydrate and lipid accumulates from 13.2% to 18.8% and 16.06% to 30.66% respectively. Under low light intensity, culture (LL) attained highest protein

content 44.73% and lowest carbohydrate and lipid content 13.2% and 16.06% respectively. The pigments content of *I. galbana* significantly ( $>0.05$ ) affected by light intensity in all cultures. With increased light intensities photosynthetic pigment chl a & chl c decreased from  $4.67 \pm 0.28$  to  $2.99 \pm 0.032$  mg/L and  $3.82 \pm 0.35$  to  $2.74 \pm 0.24$  mg/L respectively. Whereas carotenoid content increases from  $3.24 \pm 0.63$  to  $4.89 \pm 0.25$  mg/L. Similar trend is found in other studies *Dunaliella salina* [16], *Chlorella zofingiensis* [17], *Isochrysis galbana* [18].

Table 1: Effect of Light Intensity on Biochemical composition & Pigments

Biochemical Composition	Low light	Medium light	High light
Dry Weight	0.602±0.025	0.97±0.028	0.83±0.042
Protein	44.73±5.0	35.26 ±3.15	27.06± 0.35
Carbohydrate	13.2 ±.36	15.8 +0.62	18.8± 0.20
Lipid	16.06 ±0.70	23.33±0.611	30.66 ± 0.30
Chl a	4.67±0.28	3.555±0.35	2.99±0.03
Chl c	3.82±0.354	2.23±0.24	2.74±0.24
Total Carotenoid	3.24±0.636	3.35±0.12	4.89±0.256
Car/T Chl	0.38±0.03	0.578±0.56	0.853±0.17

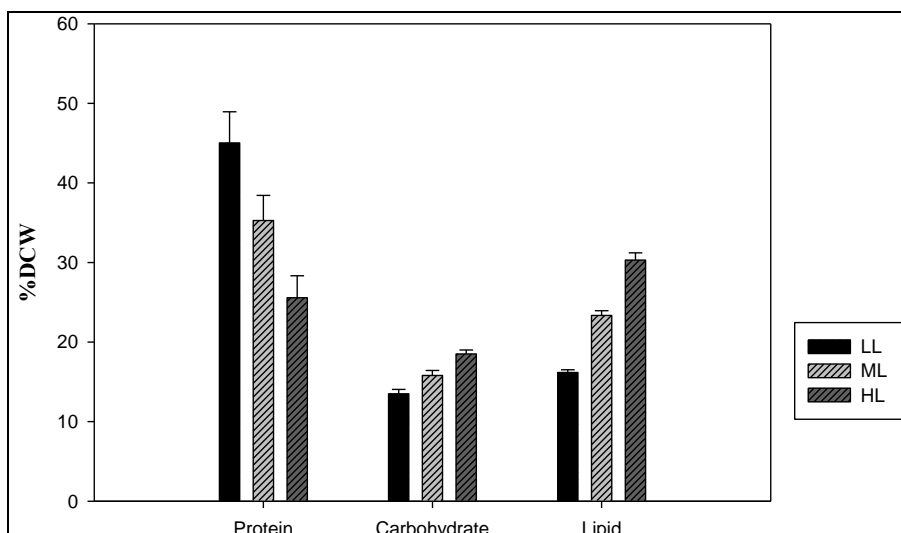
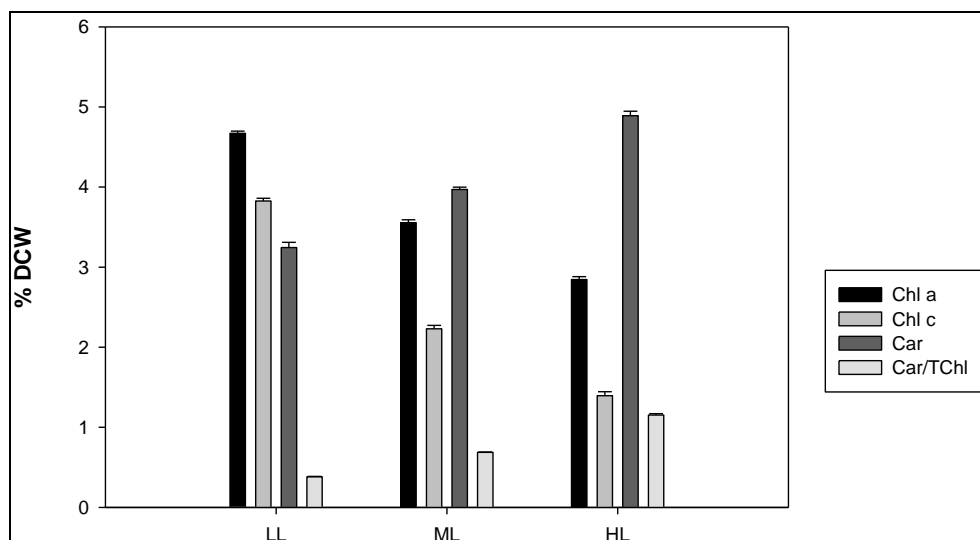


Fig 3: Biochemical variation under different light intensity



**Fig 4:** Variation of pigments under different variables

## 4. Discussion

### 4.1 Growth

Since *I. galbana* is a photosynthetic microalgae, light is determinant factor for their growth and photosynthetic activity [19, 20]. The growth curve of culture under low light intensities has shorter lag phase and longer stationary phase than other treatments. Therefore, the stationary phase may easily to achieve within its cultures. Growth pattern was investigated from optical density reveals that high light intensity stimulates the growth the *Isochrysis galbana* more efficiently than low light intensity. This could be because light is the energy source for photoautotroph's. They utilize light energy to fix carbon dioxide (CO<sub>2</sub>) to organic compounds. It also have been seen under stress such as high light intensities microalgae tend to increase in size in order to survive in stressful environments [21]. When the culture becomes very dense the growth of phytoplankton is usually slowed by light limitation [22]. Possibly excess of cells produces self shading in culture [23]. In this study *I. galbana* achieved highest growth rate under medium light intensity while further increase in light intensity lowers specific growth rate. It is in consistent with other findings in which *I. galbana* culture achieved a high growth rate at light intensities of 25–100  $\mu\text{mol m}^{-2} \text{S}^{-1}$  and light inhibitory effect occurred at 200  $\mu\text{mol photon m}^{-2} \text{s}^{-1}$  [24]. In our study, between three light intensities culture under medium light intensity of 125  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  had the highest biomass density suggesting that the optimum light regime for *Isochrysis galbana* is 125  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ . This implies that high light intensity may result in photoinhibition of *I. galbana*. This result were consistent with previous work found irradiance level 175  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  saturates photosynthesis and growth of all *Isochrysis galbana* cultures [25]. Maximum level of *Nannochloropsis sp.* growth was obtained by increasing light intensity up to 10 000 Lux (135  $\mu\text{mol photons.m}^{-2}\text{s}^{-1}$ ) which was in harmony with findings of this study (125  $\mu\text{mol photons.m}^{-2}\text{s}^{-1}$  [26]. Similar result are comparable with the finding of other groups also reported light intensities in the range of 45-120  $\mu\text{mol photons m}^{-2}\text{s}^{-1}$  saturating the growth of most of microalgae species [27]. The biomass content significantly decreased with depletion of light

from 0.97g/L to 0.602g/L. This may be due to decrease in photosynthesis, cell size and growth rates [28, 29]. The lowest dry weight have been found in cell grown in low light intensity (LL) resulted.  $602 \pm 0.062 \text{ g/L DW}$ .

### 4.2 Biochemical Composition & Pigments

Light availability is one of the chief limiting factors for growth, biomass and production of various metabolites [30]. Light quality and quantity determinates the amount of light energy available to photosynthetic organisms to conduct their metabolic activities Under high light intensity, carbohydrate and lipid in *Isochrysis galbana* accumulates significantly in order to avoid photo-oxidative damage excess light is converted to chemical energy [31, 32, 33]. The accumulation of energy-rich compounds, such as protein, lipids and carbohydrates could occur in many microalgae species under stress conditions such as light, nutrient limitation [34, 35, 36]. Stress such as high light and nitrate depletion affects the metabolic activity of cell and divert the cell from protein synthesis to carbohydrate and lipid accumulation. Lipids and carbohydrates are the preferred storage products in various stress conditions because they are hydrophobic in nature and have highly reduced states [37]. In addition they efficiently packed in small compartment of cells and can also be used during adverse conditions for cell survival and proliferation. The result of this study is in consistent with other studies reported high light intensity before photoinhibition (400nm) leads to accumulation of lipid and carbohydrates [38, 39]. Although, some studies found no significant effects of irradiances on lipid and carbohydrate accumulation [40] It have been seen under high light intensities carotenoids accumulates to prevent the absorption of surplus light energy by the photosynthetic machinery and thereby protect the photosynthetic apparatus from oxidative stress. In our study, at high light intensity, *I. galbana* synthesis less photosynthetic unit (chlorophyll a or accessory pigments) and accumulate carotenoids attributed to prevent photo damage and to boost photoprotective action.

## 5. Conclusion

The result of our study indicated that the light intensity is capable to manipulate the biochemical composition of *I. galbana*, producing either valuable proteins under low light intensity or producing carbohydrates and lipids under higher light intensities. Therefore, optimization of environmental condition enhanced the accumulation of particular metabolites which could be used for aquaculture, human nutrition, biofuel and other industrial application. On the basis of our findings we concluded that the optimum light intensity for *I. galbana* is  $125 \mu\text{mol photon m}^{-2} \text{ s}^{-1}$  shows maximum biomass dry weight. High light intensity promotes the accumulation of carotenoids, lipid and carbohydrates to protect the cell from damaging effect of high light and shifts metabolic physiology from protein synthesis to energy reserve.

## 6. Acknowledgment

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## 7. References

1. Pernet F, Tremblay R, Demers E, Roussy M. Variation of lipid class and fatty acid composition of *Chaetoceros muelleri* and *Isochrysis sp.* grown in a semi-continuous system. *Aquaculture*. 2003; 221(1):393-406.
2. Batista AP, Ambrosano L, Graca S, Sousa C, Marques PA, Ribeiro B. Combining urban wastewater treatment with biohydrogen production—An integrated microalgae-based approach. *Bioresource Technology*. 2015; 184:230-235.
3. Cai T, Park SY, Li Y. Nutrient recovery from wastewater streams by microalgae: Status and prospects. *Renewable and Sustainable Energy Reviews*. 2013; 19:360-369.
4. Song T, Martensson L, Eriksson T, Zheng W, Rasmussen U. Biodiversity and seasonal variation of the cyanobacterial assemblage in a rice paddy field in Fujian, China. *The Federation of European Microbiology Societies Microbiology Ecology*. 2005; 54:131-140.
5. Chacon-Lee TL, Gonzalez-Marino GE. Microalgae for “Healthy” Foods—Possibilities and Challenges. *Comprehensive Reviews in Food Science and Food Safety*. 2010; 9:655-675.
6. Torres FA, Passalacqua TG, Velásquez AM, de Souza RA, Colepicolo P, Graminha MA. New drugs with antiprotozoal activity from marine algae: a review. *Revista Brasileira de Farmacognosia*. 2014; 24(3):265-76.
7. Kim G, Bae J, Lee K. Nitrate repletion strategy for enhancing lipid production from marine microalga *Tetraselmis sp.* *Bioresour Technol*. 2016; 205:274-279.
8. Khoeyi ZA, Seyfabadi J, Ramezanpour Z. Effect of light intensity and photoperiod on biomass and fatty acid composition of the microalgae, *Chlorella vulgaris*. *Aquac. Int*. 2012; 20:41-49.
9. Seyfabadi J, Ramezanpour Z, Khoeyi ZA. Protein, fatty acid, and pigment content of *Chlorella vulgaris* under different light regimes *J Appl. Phycol*. 2011; 23:721-726.
10. Yu WL, Ansari W, Schoepp NG, Hannon MJ, Mayfield SP, Burkart MD. Modifications of the metabolic pathways of lipid and triacylglycerol production in microalgae. *Microbial cell factories*. 2011; 10(1):91.
11. Guillard RRL. Culture of phytoplankton for feeding marine invertebrates, edited by Smith WL, Chanie M. H (Plenum Press, New York), 1975, 29-60.
12. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin Phenol reagent *J Biol. Chem*, 1951; 193:265-275.
13. DuBois M, Gilles KA, Hamilton JK, Rebers PT, Smith F. Colorimetric method for determination of sugars and related substances. *Analytical chemistry*. 1956; 28(3):350-6.
14. Bligh EG, Dyer WJ. A rapid method of total lipid extraction and purification. *Canadian Biochem. Physiol*. 1959; 37:911-917.
15. Jeffrey ST, Humphrey GF. New spectrophotometric equations for determining chlorophylls a, b, c 1 and c 2 in higher plants, algae and natural phytoplankton. *Biochemie und Physiologie der Pflanzen*. 1975; 167(2):191-4.
16. Lamers PP, Janssen M, De Vos RCH, Bino RJ, Wijffels RH. Exploring and exploiting carotenoid accumulation in *Dunaliella salina* for cell-factory applications. *Trends Biotechnol*. 2008; 26:631-638.
17. Mulders KJM, Lamers PP, Wijffels RH, Martens DE, Dynamics of biomass composition and growth during recovery of nitrogen-starved *Chromochloris zofingiensis*. *Appl Microbiol Biotechnol*. 2015; 99:1873-1884.
18. Roopnarain A, Gray VM, Sym S. Influence of nitrogen stress on *Isochrysis galbana* strain U4, a candidate for biodiesel production, *Phycol. Res*. 2014; 62(4):237-249.
19. Wahidin S, Idris A, Shaleh SRM. The influence of light intensity and photoperiod on the growth and lipid content of microalgae *Nannochloropsis sp.*, *Bioresour. Technol*. 2013; 129:7-11.
20. Wang L. Research on the Relevant Factors of the Algal Growth in Hydrodynamics Condition. Chongqing University, 2006.
21. Leal S, Alejandra-Medina M, Alejandro-Guerrero M, Piña P, Nieves M, Curbelo R. Concentración y composiciones orgánica y proximal de dos especies de diatomeas bentónicas a diferentes salinidades, *Universidad y ciencia*. 2013; 29(1):45-52.
22. Harrison PJ, Thompson PA, Calderwood GS. Effects of nutrient and light limitation on the biochemical composition of phytoplankton, *Journal of Applied Phycology*. 1990; 2:45-56.
23. Johnsen G, Sakshaug E. Biooptical characteristics of PSII and PSI in 33 species (13 pigment groups) of marine phytoplankton, and the relevance for pulse-amplitude-modulated and fast-repetition-rate fluorometry. *Journal of Phycology*. 2007; 43(6):1236-51.
24. Alkhamis Y, Qin JG. Cultivation of *Isochrysis galbana* in phototrophic, heterotrophic, and mixotrophic conditions. *BioMed research international*, 2013; 10, 2013.
25. Herrig R, Falkowsk PG. Nitrogen Limitation in *Isochrysis Galbana* (Haptophyceae): Photosynthetic Energy Conversion and Growth Efficiencies, *Journal of Phycology*. 1989; 3(25):462-471.

26. Cheirsilp B, Torpee S. Enhanced growth and lipid production of microalgae under mixotrophic culture condition: effect of light intensity, glucose concentration and fed-batch cultivation. *Bioresour Technol.* 2012; 110:510-6.
27. Chan TA. Comparative physiological study of marine diatoms and dinoflagellates in relation to irradiance and cell size under continuous light *J Phycol.* 1978; 14:396-402.
28. Simionato D, Block MA, La Rocca N, Jouhet J, Maréchal E, Finazzi G, *et al.* The response of *Nannochloropsis gaditana* to nitrogen starvation includes de novo biosynthesis of triacylglycerols, a decrease of chloroplast galactolipids, and reorganization of the photosynthetic apparatus. *Eukaryot. Cell.* 2013; 12:665-676.
29. Li Y, Horsman M, Wang B, Wu N, Lan CQ. Effect of nitrogen source on cell growth and lipid accumulation of green alga *Neochloris oleoabundans*. *Appl. Microbiol. Biot.* 2008; 81:629-636.
30. Cordero BF, Couso I, León R, Rodríguez H, Vargas MÁ. Enhancement of carotenoids biosynthesis in *Chlamydomonas reinhardtii* by nuclear transformation using a phytoene synthase gene isolated from *Chlorella zofingiensis*. *Applied microbiology and biotechnology.* 2011; 91(2):341-51.
31. Asada K. Production and action of active oxygen species in photosynthetic tissues, edited by Foyer CH, Mullineaux PM (CRC, Boca Raton). 1994, 77-104.
32. Rabbani S, Beyer P, Lintig J, Hugueney P, Kleinig H. Induced  $\beta$ -carotene synthesis driven by triacylglycerol deposition in the unicellular alga *Dunaliella bardawii*. *Plant Physiol.* 1998; 116:1239-1248.
33. Mendoza H, Martel A, Jimenez del Rio M, Garcia Reina G. Oleic acid is the main fatty acid related with carotenogenesis in *Dunaliella salina* *J Appl Phycol.* 1999; 11:15-19.
34. Niyogi K. Photoprotection revisited: genetic and molecular approaches. *Annu Rev Plant Physiol Mol Biol.* 1999; 50:333-359.
35. Rao AR, Dayananda C, Sarada R, Shamala TR, Ravishankar GA. Effect of salinity on growth of green alga *Botryococcus braunii* and its constituents. *Bioresour. Technol.* 2007; 98:560-564.
36. Li Y, Horsman M, Wang B, Wu N, Lan CQ. Effects of nitrogen sources on cell growth and lipid accumulation of green alga *Neochloris oleoabundans*. *Appl. Microbiol. Biotechnol.* 2008; 81:629-636.
37. Courchesne NMD, Parisien A, Wang B, Lan CQ. Enhancement of lipid production using biochemical, genetic and transcription factor engineering approaches *J Biotechnol.* 2009; 141(1-2):31-41.
38. Sandnes J, Källqvist T, Wenner D, Gislerød HR. Combined influence of light and temperature on growth rates of *Nannochloropsis oceanic* linking cellular responses to large-scale biomass production. *Journal of Applied Phycology.* 2005; 17:515-525.
39. Sun Yingying, Changhai Wang. The optimal growth conditions for the biomass production of *Isochrysis galbana* and the effects that phosphorus,  $Zn^{2+}$ ,  $CO_2$ , and light intensity have on the biochemical composition of *Isochrysis galbana* and the activity of extracellular CA, *Biotechnology and Bioprocess Engineering.* 2009; 14(2):225-231
40. Carvalho AP, Monteiro CM, Malcata FX. Simultaneous effect of irradiance and temperature on biochemical composition of the microalga *Pavlova lutheri*, *Journal of Applied Phycology.* 2009; 21 (5):543-552.